Cationic Ruthenium Catalysts for Alkyne Annulations with Oximes by C–H/N–O Functionalizations

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Supporting Information

ABSTRACT: Cationic ruthenium(II) complexes enabled efficient redox-neutral annulations of alkynes with readily available oximes. The catalytic dehydrative C-H/N-O bond functionalization proved to be broadly applicable and was shown to proceed through a reversible cyclometalation.



INTRODUCTION

The modular assembly of regioselectively decorated heterocycles is of central importance because of their prevalence in natural products and drugs.¹ Particularly, isoquinolines are key building blocks in organic synthesis and represent structural motifs of alkaloids with, among others, cardiovascular, antiinflammatory or antimalarial bioactivities.² As a result, there is a continued strong demand for versatile isoquinoline syntheses. While transition metal-catalyzed annulations with prefunctionalized *ortho*-halo-substituted benzimines were previously developed,³ a more atom- and step-economical strategy was realized with methods that capitalize upon $C-H^4$ bond activation. Hence, the research groups of Jun, Bergman and Ellman, Cheng, Fagnou, Satoh and Miura, Chiba, Rovis as well as Li exploited chelation assistance for rhodium-catalyzed⁵ isoquinoline and pyridine syntheses.⁶

In 2011, we reported on the use of less-expensive ruthenium⁷ complexes for oxidative annulations of alkynes by benzamides through C–H/N–H bond cleavages.⁸ Notably, this approach proved more generally applicable.⁹ Indeed, ruthenium(II) biscarboxylates^{4b} also set the stage for redox-neutral dehydrative annulations of alkynes with free hydroxamic acids through C–H/N–O functionalizations to furnish isoquinolones (Scheme 1a).¹⁰

In consideration of the important bioactivities of substituted isoquinolines,^{1–3} we consequently became attracted by utilizing ruthenium(II) complexes for step-economical isoquinoline syntheses in the absence of external oxidants (Scheme 1b), the results of which we report herein. In contrast to previous studies,^{9,10} we observed a remarkably high catalytic activity of carboxylate-free¹¹ cationic ruthenium(II) catalysts and obtained detailed mechanistic insight on reversible cyclometalations in dehydrative isoquinoline syntheses.

RESULTS AND DISCUSSION

We initiated our studies by exploring representative metal carboxylates and solvents for the envisioned dehydrative annulation of alkyne 2a by oxime 1a (Table 1). Among various cocatalytic additives, KPF_6 gave rise to the most satisfactory results (entries 1–9),¹² while MeOH proved to be

Scheme 1. Ruthenium-Catalyzed C-H/N-O Functionalizations



the solvent of choice for the dehydrative annulation (entries 9–12). However, $[RuBr_2(p\text{-cymene})]_2$ delivered the desired isoquinoline **3aa** in a reduced yield as compared to a reaction with $[RuCl_2(p\text{-cymene})]_2$ as the catalyst (entries 9, and 13). The high catalytic activity exerted by the KPF₆-derived ruthenium catalyst was suggestive of an in situ formation of a cationic ruthenium(II) species (vide infra).

Subsequently, we explored the scope of the optimized cationic ruthenium(II) complex in the redox-neutral annulation of alkynes 2 (Scheme 2). Thus, a variety of differently substituted isoquinolines 3ba-3ea bearing valuable electrophilic functional groups was chemoselectively accessed. Furthermore, oximes 1 bearing sterically more demanding substituents furnished the desired products 3fa-3ma, as did bicyclic substrate 1n. The catalytic system was not restricted to tolane (2a) but allowed for the efficient conversion of different aryl- and alkyl-substituted alkynes 2b-2f as well. Intramolecular competition experiments with *meta*-substituted arenes 10 and 1p revealed a significant change in site-selectivity. Thus, isoquinoline 3oa was generated through steric interactions,

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Table 1. Optimization of C-H/N-O Functionalization^a



^{*a*}Reaction conditions: **1a** (0.50 mmol), **2a** (1.00 mmol), $[RuCl_2(p-cymene)]_2$ (5.0 mol %), additive (30 mol %), solvent (2 mL), 60 °C, isolated yields. ^{*b*}GC conversion. ^{*c*}4 Å mol. sieves (100 mg). ^{*d*}[RuBr₂(*p*-cymene)]₂ (5.0 mol %).





^{*a*}In the presence of 4 Å mol. sieves.

while oxime 1p displaying a secondary directing group was functionalized at the sterically more congested C-2 position.

Importantly, intermolecular dehydrative annulations with challenging unsymmetrical alkynes 2 occurred with excellent regioselectivities (Scheme 3).¹³ Hence, the cationic ruthenium-





(II) complex furnished the 3-aryl-substituted isoquinolines **3ag–3aq** with high yields. Moreover, the catalyst proved tolerant of useful functional groups, such as esters, ketones, free hydroxyl groups as well as aryl and alkyl halides.

Given the remarkable efficacy of the in situ generated ruthenium(II) catalyst, we subsequently probed the performance of preformed cationic complex $[Ru_2Cl_3(p-cymene)_2][PF_6]$ (4)^{12,14} (Scheme 4). Importantly, well-defined complex 4 displayed an activity as well as chemo-, regio- and site-selectivity being comparable to the one displayed by the in situ formed catalytic system.

Mechanistic Studies. Considering the high catalytic efficacy exerted by the optimized cationic catalyst, we became interested in delineating its mode of action. To this end, we performed intermolecular competition experiments between different alkynes 2, which revealed electron-deficient substrates to be more reactive (Scheme 5).

Moreover, intermolecular competition experiments with *para*-substituted oximes **1** highlighted electron-rich arenes to be preferentially converted (Scheme 6).

Mechanistic studies with deuterated solvent MeOH- d_4 and isotopically labeled substrate **1a**- d_5 showed a significant H/D scrambling, albeit solely occurring in the presence of an alkyne (Scheme 7). These observations were, hence, indicative of a reversible C–H bond metalation step by an alkyne-coordinated ruthenium complex, likely proceeding through base assistance. It is further noteworthy that deuterium was also incorporated into the methyl^{6f,15} group of product **3ae**- d_n when using MeOH- d_4 as the solvent (Scheme 7b).

On the basis of these mechanistic studies, we propose a catalytic cycle to involve an initial reversible base-assisted cycloruthenation with an alkyne-coordinated cationic complex, furnishing metalacycle 5, presumably, via a S_EAr -type mechanism (Scheme 8). Subsequent migratory insertion of

Scheme 4. Well-Defined Cationic Complex 4 as the Catalyst



Scheme 5. Competition Experiment with Alkynes 2



the alkyne 2 generates key intermediate 6, while reductive elimination delivers product 3. Finally, reoxidation by the internal oxidant via N-O bond cleavage regenerates the catalytically active species.

CONCLUSIONS

In summary, we have reported on ruthenium-catalyzed redoxneutral annulations of alkynes by oximes through C–H/N–O functionalizations. Contrary to previous studies, ^{9,10} carboxylatefree cationic ruthenium(II) catalysts were found to be most effective, which allowed for an expedient isoquinoline synthesis via reversible cyclometalation.

EXPERIMENTAL SECTION

General Remarks. Catalytic reactions were carried out under a N_2 atmosphere using predried glassware. H_2O was degassed and purged with N_2 , DMF was dried and distilled over CaH₂, *t*-AmOH was dried and distilled over sodium, and MeOH was dried and distilled over Mg(OMe)₂. All ketoximes 1 were synthesized according to a previously described procedure.¹⁶ Internal alkynes 2 were prepared

Scheme 6. Intermolecular Competition Experiments







according to previously described methods and were purified via Kugelrohr distillation.^{17,18} [Ru₂Cl₃(*p*-cymene)₂][PF₆] (4) was synthesized according to a previously reported procedure.¹² Other chemicals were obtained from commercial sources and were used without further purification. Yields refer to isolated compounds, estimated to be >95% pure as determined by ¹H NMR and GC analysis. NMR spectra were recorded in the solvent indicated; chemical shifts (δ) are given in ppm. High resolution mass spectrometry (HRMS): FTICR.

Representative Procedure for the Ruthenium-Catalyzed Synthesis of Isochinolines via Direct C–H/N–O Bond Functionalization, Synthesis of 1-Methyl-3,4-diphenylisoquinoline (3aa). A mixture of acetophenone oxime (1a) (68 mg, 0.50 mmol), diphenylacetylene (2a) (178 mg, 1.00 mmol), $[RuCl_2(p-cymene)]_2$ (15.3 mg, 0.025 mmol, 5.0 mol %), and KPF₆ (28 mg, 0.15 mmol, 30 mol %) in MeOH (2.00 mL) was stirred at 60 °C for 24 h. At ambient temperature, EtOAc (15 mL) was added, and the solvents



were removed in vacuo. Purification of the remaining residue by column chromatography (*n*-hexane/EtOAc: 10/1) yielded **3aa** (119 mg, 81%) as a white solid (mp 152–155 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.23–8.20 (m, 1H), 7.71–7.67 (m, 1H), 7.63–7.57 (m, 2H), 7.44–7.32 (m, SH), 7.29–7.17 (m, SH), 3.11 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 157.7 (C_q), 149.4 (C_q), 141.0 (C_q), 137.6 (C_q), 136.0 (C_q), 131.4 (CH), 130.2 (CH), 129.9 (CH), 129.1 (C_q), 128.1 (CH), 126.6 (CH), 127.1 (CH), 126.9 (CH), 126.5 (CH), 126.2 (CH), 126.1 (C_q), 125.5 (CH), 22.7 (CH₃); IR (ATR) 3025, 1567, 1389, 1334, 1072, 1026, 765, 695, 612, 563, 496 cm⁻¹; MS (EI) *m/z* (relative intensity) 295 (50) [M⁺], 294 (100), 278 (5), 252 (17), 177 (15), 146 (6), 43 (14); HR-MS (EI) *m/z* calcd for C₂₂H₁₇N 295.1361, found 295.1348. The spectral data are in accordance with those reported in the literature.¹⁹

1,6-Dimethyl-3,4-diphenylisoquinoline (3ba). The representative procedure was followed using 1b (75 mg, 0.50 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 10/1) yielded 3ba (131 mg, 85%) as a white solid (mp 160–163 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.09 (d, J = 9.2 Hz, 1H), 7.44-7.39 (m, 2H), 7.39-7.30 (m, 5H), 7.25-7.14 (m, 5H), 3.05 (s, 3H), 2.43 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 157.3 (C_q), 149.5 (C_q), 141.1 (C_q), 140.1 (C_q), 137.7 (C_a), 136.2 (C_a), 131.4 (CH), 130.2 (CH), 128.7 (C_a), 128.6 (CH), 128.1 (CH), 127.5 (CH), 127.0 (CH), 126.8 (CH), 125.4 (CH), 125.0 (CH), 124.5 (C_a), 22.6 (CH₃), 22.1 (CH₃); IR (ATR) 3062, 1495, 1444, 1385, 1336, 1071, 1029, 813, 767, 755, 696, 614 cm⁻¹; MS (EI) m/z (relative intensity) 309 (40) [M⁺], 308 (100), 293 (5), 265 (5), 252 (12), 146 (5), 43 (4); HR-MS (ESI) m/z calcd for $C_{23}H_{19}N$ + H⁺ 310.1590, found 310.1592. The spectral data are in accordance with those reported in the literature.

6-(Trifluoromethyl)-1-methyl-3,4-diphenylisoquinoline (**3ca**). The representative procedure was followed using 1c (102 mg, 0.50 mmol) and diphenylacetylene (**2a**) (178 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 12/1) yielded **3ca** (118 mg, 65%) as an orange solid (mp 109–114 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.31 (d, *J* = 8.8 Hz, 1H), 7.96 (d, *J* = 1.8 Hz, 1H), 7.75 (dd, *J* = 8.8, 1.8 Hz, 1H), 7.40–7.33 (m, 5H), 7.23–7.16 (m, 5H), 3.10 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 157.8 (C_q), 150.9 (C_q), 140.4 (C_q), 136.5 (C_q), 135.4 (C_q), 131.5 (q, ²*J*_{C-F} = 32 Hz, C_q), 131.2 (CH), 130.2 (CH), 129.7 (C_q), 128.5 (CH), 127.7 (CH), 127.6 (CH), 127.3 (CH), 127.0 (C_q), 126.8 (CH), 123.9 (q, ³*J*_{C-F} = 5 Hz, CH), 123.8 (d, ¹*J*_{C-F} = 272 Hz, C_q), 122.2 (q, ³*J*_{C-F} = 3 Hz, CH), 22.8 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -62.9 (s); IR (ATR) 2958, 1555, 1336, 1305, 1257, 1176, 1155, 1134, 1082, 909, 769, 696, 618 cm⁻¹; MS (EI) *m/z* (relative intensity) 363 (50) [M⁺], 362 (100), 252 (8), 146 (5), 43 (5); HR-MS (EI) *m/z* calcd for $C_{23}H_{16}F_{3}N$ 363.1235, found 363.1219. The spectral data are in accordance with those reported in the literature. 20

6-Bromo-1-methyl-3,4-diphenylisoquinoline (3da). The representative procedure was followed using 1d (107 mg, 0.50 mmol) diphenylacetylene (2a) (178 mg, 1.00 mmol) and molecular sieves 4 Å (100 mg). Purification by column chromatography (*n*-hexane/EtOAc: 20/1) yielded 3da (102 mg, 54%) as a yellow solid (mp 193-195 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.06 (d, J = 8.9 Hz, 1H), 7.80 (d, J = 2.0 Hz, 1H), 7.67 (dd, J = 8.9, 2.0 Hz, 1H), 7.39-7.30 (m, 5H), 7.23-7.14 (m, 5H), 3.06 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 157.6 (C_q), 150.5 (C_q), 140.5 (C_q), 137.3 (C_q), 136.7 (C_q), 131.2 (CH), 130.0 (CH), 129.9 (CH), 128.3 (CH), 128.3 (CH), 128.2 (C₀), 127.6 (CH), 127.4 (CH), 127.2 (CH), 127.1 (CH), 125.0 (C_a), 124.5 (C_a), 22.8 (CH₃); IR (ATR) 3064, 1597, 1561, 1481, 1445, 1386, 1329, 1259, 1071, 1029, 751, 697 cm⁻¹; MS (EI) *m/z* (relative intensity) 374 (100) [M + H⁺], 293 (26), 252 (28), 189 (15), 146 (14), 43 (56); HR-MS (ESI) m/z calcd for $C_{22}H_{16}BrN + H^+$ 374.0539, found 374.0538. The spectral data are in accordance with those reported in the literature.²⁰

1-Methyl-3,4,6-triphenylisoquinoline (3ea). The representative procedure was followed using 1e (106 mg, 0.50 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 15/1) yielded 3ea (101 mg, 54%) as a pale orange solid (mp 176-178 °C): ¹H NMR (300 MHz, $CDCl_3$) $\delta = 8.28$ (dd, J = 8.2, 1.1 Hz, 1H), 7.87 (s, 1H), 7.86 (d, J =8.2, 1.8 Hz 1H), 7.62-7.53 (m, 2H), 7.49-7.14 (m, 14H), 3.11 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 157.5 (C_a), 150.0 (C_a), 142.5 (C_q), 141.0 (C_q), 140.4 (C_q), 137.5 (C_q), 136.3 (C_q), 131.4 (CH), 130.2 (CH), 129.3 (C_q), 128.9 (CH), 128.2 (CH) 127.9 (CH), 127.6 (CH), 127.5 (CH), 127.2 (CH), 126.9 (CH), 126.2 (CH), 126.2 (CH), 125.2 (C_a), 124.0 (CH), 22.7 (CH₃); IR (ATR) 3058, 1611, 1567, 1434, 1339, 955, 893, 831, 760, 751, 690, 611 cm⁻¹; MS (EI) *m*/ z (relative intensity) 371 (68) [M⁺], 370 (100), 354 (3), 327 (5), 292 (2), 252 (4), 77 (3); HR-MS (EI) *m/z* calcd for C₂₈H₂₁N 371.1674, found 371.1657. The spectral data are in accordance with those reported in the literature.²⁰

1-Ethyl-3,4-diphenylisoquinoline (3fa). Representative procedure A was followed using 1f (75 mg, 0.50 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 15/1) yielded 3fa (141 mg, 91%) as a pale brown solid (mp 113-115 °C): ¹H NMR (300 MHz, $CDCl_3$) δ = 8.29–8.20 (m, 1H), 7.70–7.62 (m, 1H), 7.61–7.50 (m, 2H), 7.42-7.30 (m, 5H), 7.26-7.12 (m, 5H), 3.44 (q, J = 7.6 Hz, 2H), 1.53 (t, J = 7.6 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) $\delta = 162.2$ (C_q), 149.2 (C_q), 141.1 (C_q), 137.7 (C_q), 136.3 (C_q), 131.4 (CH), 130.3 (CH), 129.7 (CH), 128.9 (C_a), 128.2 (CH), 127.5 (CH), 127.1 (CH), 126.8 (CH), 126.4 (CH), 126.4 (CH), 125.3 (C_a), 125.1 (CH), 28.8 (CH₂), 13.9 (CH₃); IR (ATR) 2935, 1552, 1445, 1382, 1260, 1072, 1028, 753, 694, 607, 561 cm⁻¹; MS (EI) m/z (relative intensity) 309(55) [M⁺], 308(100), 293(14), 280(8), 252(5), 146(5), 69(6); HR-MS (EI) m/z calcd for C₂₃H₁₉N 309.1517, found 309.1505. The spectral data are in accordance with those reported in the literature.19

1-Ethyl-6-methoxy-3,4-diphenylisoquinoline (3ga). The representative procedure was followed using 1g (94 mg, 0.52 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 12/1) yielded 3ga (168 mg, 95%) as a pale yellow solid (mp 158-161 °C): ¹H NMR (300 MHz, $CDCl_3$) $\delta = 8.14$ (d, J = 9.2 Hz, 1H), 7.40–7.28 (m, 5H), 7.26–7.11 (m, 6H), 6.91 (d, J = 2.6 Hz, 1H), 3.71 (s, 3H), 3.37 (q, J = 7.6 Hz, 2H), 1.50 (t, J = 7.6 Hz, 3H); ^{13}C NMR (75 MHz, CDCl₃) δ = 161.6 (C_q) , 160.3 (C_q) , 150.0 (C_q) , 141.3 (C_q) , 138.4 (C_q) , 138.0 (C_q) , 131.3 (CH), 130.3 (CH), 128.3 (C_q), 128.2 (CH), 127.5 (CH), 127.0 (CH), 127.0 (CH), 126.8 (CH), 121.0 (C_q), 119.0 (CH), 104.6 (CH), 55.1 (CH₃), 28.8 (CH₂), 14.1 (CH₃); IR (ATR) 2970, 1617, 1574, 1454, 1411, 1232, 1146, 1018, 757, 699, 662, 527 cm⁻¹; MS (EI) m/z(relative intensity) 339 (55) [M⁺], 338 (100), 323 (5), 295 (10), 280 (12), 267 (7), 239 (8); HR-MS (EI) m/z calcd for C₂₄H₂₁NO 339.1623, found 339.1613.

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1-Ethyl-6-fluoro-3,4-diphenylisoquinoline (3ha). The representative procedure was followed using **1h** (83 mg, 0.50 mmol) and diphenyl acetylene (**2a**) (178 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 15/1 → 12/1) yielded **3ha** (94 mg, 57%) as a white solid (mp 141–142 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.27 (dd, *J* = 9.2, 5.7 Hz, 1H), 7.45–7.10 (m, 12H), 3.42 (q, *J* = 7.5 Hz, 2H), 1.53 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 163.0 (d, ¹*J*_{C-F} = 245 Hz, C_q), 162.0 (C_q), 150.7 (C_q), 140.8 (C_q), 138.4 (d, ³*J*_{C-F} = 10 Hz, C_q), 137.3 (C_q), 131.2 (CH), 130.3 (CH), 128.7 (d, ⁴*J*_{C-F} = 5 Hz, C_q), 128.4 (CH), 128.2 (d, ³*J*_{C-F} = 10 Hz, CH), 127.6 (CH), 127.1 (CH), 122.5 (d, ⁴*J*_{C-F} = 1 Hz, C_q), 116.6 (d, ²*J*_{C-F} = 25 Hz, CH), 110.0 (d, ²*J*_{C-F} = 22 Hz, CH), 28.9 (CH₂), 13.9 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -107.9 (s); IR (ATR) 2973, 1619, 1573, 1447, 1386, 1182, 1072, 876, 788, 753, 697 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 327 (53) [M⁺], 326 (100), 311 (12), 298 (10), 98 (10), 74 (6), 57 (10), 43 (20); HR-MS (ESI) *m*/*z* calcd for C₂₃H₁₈FN + H⁺ 328.1496, found 328.1498.

3,4-Diphenyl-1-n-propylisoquinoline (3ia). The representative procedure was followed using 1i (82 mg, 0.50 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 50/1) yielded 3ia (144 mg, 89%) as a yellow solid (mp 118–120 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.23-8.14 (m, 1H), 7.64-7.56 (m, 1H), 7.54-7.45 (m, 2H), 7.37-7.22 (m, 5H), 7.20-7.05 (m, 5H), 3.32 (t, J = 7.7 Hz, 2H), 1.95 (dt, J = 7.7, 7.3 Hz 2H), 1.08 (t, J = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl_3 $\delta = 161.3 (C_q), 149.3 (C_q), 141.1 (C_a), 137.8 (C_a), 136.3$ (C_q), 131.4 (CH), 130.3 (CH), 129.6 (CH), 128.9 (C_q), 128.2 (CH), 127.5 (CH), 127.5 (CH), 127.1 (CH), 126.9 (CH), 126.3 (CH), 125.6 (C_q), 125.2 (CH), 37.7 (CH₂), 23.2 (CH₂), 14.5 (CH₃); IR (ATR) 3063, 2962, 1612, 1568, 1550, 1444, 1385, 1087, 1031, 757, 697 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 323 (22) [M⁺], 295 (100) 308 (16), 252 (7); HR-MS (EI) m/z calcd for $C_{24}H_{21}N - H^+$ 322.1596, found 322.1607.

1-n-Butyl-3,4-diphenylisoquinoline (3ja). The representative procedure was followed using 1j (89 mg, 0.50 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 40/1) yielded 3ja (151 mg, 89%) as a yellow solid (mp 78–80 °C): ¹H NMR (300 MHz, CDCl₃) $\delta =$ 8.27-8.21 (m, 1H), 7.69-7.64 (m, 1H), 7.60-7.54 (m, 2H), 7.40-7.31 (m, 5H), 7.26–7.15 (m, 5H), 3.42 (t, J = 8.0 Hz, 2H), 2.02–1.90 (m, 2H), 1.58 (dt, J = 7.7, 7.3 Hz, 2H), 1.04 (t, J = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 161.5 (C_q), 149.3 (C_q), 141.1 (C_q), 137.7 (C_a), 136.3 (C_a), 131.4 (CH), 130.3 (CH), 129.6 (CH), 128.8 (C_a), 128.2 (CH), 127.5 (CH), 127.5 (CH), 127.0 (CH), 126.8 (CH), 126.3 (CH), 125.4 (C_g), 125.2 (CH), 35.5 (CH₂), 32.1 (CH₂), 23.1 (CH₂), 14.1 (CH₃); IR (ATR) 3061, 2958, 2876, 1611, 1504, 1442, 1382, 1337, 1172, 1073, 763, 697 cm⁻¹; MS (EI) *m/z* (relative intensity) 337 (6) [M⁺], 295 (100), 308 (10), 252 (6); HR-MS (EI) m/z calcd for C₂₅H₂₃N - H⁺ 336.1752, found 336.1747. The spectral data are in accordance with those reported in the literature.²¹

1-*n***-Pentyl-3,4-diphenylisoquinoline (3ka).** The representative procedure was followed using **1k** (96 mg, 0.50 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 50/1) yielded **3ka** (168 mg, 96%) as a yellow oil: ¹H NMR (300 MHz, CDCl₃) δ = 8.31–8.19 (m, 1H), 7.71–7.63 (m, 1H), 7.61–7.53 (m, 2H), 7.42–7.28 (m, 5H), 7.27–7.12 (m, 5H), 3.40 (t, *J* = 7.5 Hz, 2H), 2.07–1.89 (m, 2H), 1.66–1.37 (m, 4H), 0.96 (t, *J* = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 161.5 (C_q), 149.3 (C_q), 141.1 (C_q), 137.7 (C_q), 136.3 (C_q), 131.4 (CH), 130.3 (CH), 129.6 (CH), 128.9 (C_q), 128.2 (CH), 127.5 (CH), 127.5 (CH), 127.1 (CH), 126.8 (CH), 126.3 (CH), 125.5 (C_q), 125.2 (CH), 35.8 (CH₂), 32.2 (CH₂), 29.7 (CH₂), 22.6 (CH₂), 14.1 (CH₃); IR (ATR) 3058, 2927, 2858, 1613, 1551, 1504, 1443, 1383, 1073, 1030, 763, 696, 612, 566 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 351 (5) [M⁺], 322 (10), 308 (12), 295 (100), 252 (5), 216 (5); HR-MS (EI) *m*/*z* calcd for C₂₆H₂₅N 351.1987, found 351.1998.

1,3,4-Triphenylisoquinoline (3la). The representative procedure was followed using **11** (99 mg, 0.50 mmol) and diphenylacetylene (**21**) (178 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 25/1) yielded **3la** (147 mg, 82%) as a pale yellow

solid (mp 181–184 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.20 (dm, *J* = 8.5 Hz, 1H), 7.84 (dd, *J* = 8.0, 1.6 Hz, 2H), 7.74 (dm, *J* = 8.7 Hz, 1H), 7.67–7.10 (m, 15H); ¹³C NMR (75 MHz, CDCl₃) δ = 159.9 (C_q), 149.6 (C_q), 140.9 (C_q), 139.8 (C_q), 137.5 (C_q), 136.9 (C_q), 131.3 (CH), 130.4 (CH), 130.2 (CH), 129.9 (CH), 129.7 (C_q), 128.5 (CH), 128.3 (CH), 126.6 (CH), 127.5 (CH), 127.5 (CH), 127.3 (CH), 127.0 (CH), 126.6 (CH), 126.0 (CH), 125.4 (C_q); IR (ATR) 3054, 1540, 1494, 1442, 1384, 1336, 1073, 1030, 980, 761, 700, 668, 633, 567 cm⁻¹; MS (EI) *m/z* (relative intensity) 357 (50) [M⁺], 356 (100), 278 (11), 252 (10), 177 (5); HR-MS (EI) *m/z* calcd for C₂₇H₁₉N 357.1517, found 357.1493. The spectral data are in accordance with those reported in the literature.¹⁹

1-Cyclopropyl-3,4-diphenylisoquinoline (3ma). The representative procedure was followed using 1m (81 mg, 0.50 mmol), diphenylacetylene (2a) (178 mg, 1.00 mmol) and molecular sieves 4 Å (100 mg). Purification by column chromatography (*n*-hexane/EtOAc: 200/1) yielded 3ma (135 mg, 84%) as a yellow solid (mp 148-150 °C): ¹H NMR (300 MHz, \tilde{CDCl}_3) δ = 8.50 (dd, J = 7.2, 2.0 Hz, 1H), 7.69-7.62 (m, 1H), 7.62-7.51 (m, 2H), 7.42-7.27 (m, 5H), 7.27-7.21 (m, 2H), 7.20-7.08 (m, 3H), 2.89-2.74 (m, 1H), 1.44-1.32 (m, 2H), 1.18–1.05 (m, 2H); ¹³C NMR (75 MHz, $CDCl_3$) δ = 160.6 (C_a), 148.7 (C_a), 141.2 (C_a), 138.0 (C_a), 136.1 (C_q), 131.4 (CH), 130.4 (CH), 129.6 (CH), 128.2 (CH), 128.0 (C_a), 127.3 (CH), 127.0 (CH), 126.8 (CH), 126.3 (CH), 126.3 (C_q), 126.2 (CH), 124.8 (CH), 13.6 (CH), 9.4 (CH₂); IR (ATR) 3055, 1611, 1568, 1547, 1444, 1410, 1318, 1261, 1075, 1014, 767, 694 cm⁻¹; MS (EI) m/z (relative intensity) 321 (68) [M⁺], 320 (100), 278 (5), 243 (8), 152(5), 43 (13); HR-MS (EI) m/z calcd for $C_{24}H_{19}N - H^+$ 320.1439, found 320.1438. The spectral data are in accordance with those reported in the literature. $^{\rm 22}$

9-Methyl-2,3-diphenyl-8,9-dihydro-7H-benzo[de]quinoline (3na). The representative procedure was followed using 1n (88 mg, 0.50 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 40/1) yielded 3na (157 mg, 93%) as a yellow solid (mp 145-147 °C): ¹H NMR (300 MHz, CDCl₃) δ = 7.51–7.46 (m, 2H), 7.45–7.28 (m, 7H), 7.25-7.12 (m, 4H), 3.54-3.41 (m, 1H), 3.36-3.24 (m, 1H), 3.23-3.11 (m, 1H), 2.43-2.31 (m, 1H), 2.12-1.99 (m, 1H), 1.63-1.56 (d, J = 7.1, 3H); ¹³C NMR (75 MHz, C₂D₂Cl₄, 100 °C) $\delta = 162.0$ (C_q) , 148.7 (C_q) , 141.3 (C_q) , 138.0 (C_q) , 138.0 (C_q) , 136.2 (C_q) , 131.2 (CH), 130.3 (CH), 129.3 (CH), 128.4 (C_q), 127.9 (CH), 127.0 (CH), 126.7 (CH), 126.4 (CH), 124.3 (CH), 123.3 (CH), 123.0 (C_a), 37.4 (CH), 30.8 (CH₂), 27.8 (CH₂), 19.6 (CH₃); IR (ATR) 2929, 1604, 1577, 1442, 1377, 1306, 1178, 1071, 1023, 767, 699 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 335 (91) [M⁺], 334 (100), 320 (50), 241 (8), 43 (13); HR-MS (EI) m/z calcd for $C_{25}H_{21}N - H^+$ 334.1596, found 334.1608.

3,4-Bis(4-methoxyphenyl)-1-methylisoquinoline (3ab). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 1,2-bis(4-methoxyphenyl)acetylene (2b) (238 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: $10/1 \rightarrow 8/$ 1) yielded 3ab (95 mg, 53%) as an orange solid (mp 106-110 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.20–8.11 (m, 1H), 7.73–7.62 (m, 1H), 7.60–7.49 (m, 2H), 7.32 (d, J = 8.9 Hz, 2H), 7.13 (d, J = 8.8 Hz, 2H), 6.90 (d, J = 8.8 Hz, 2H), 6.74 (d, J = 8.9 Hz, 2H), 3.83 (s, 3H), 3.75 (s, 3H), 3.04 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 158.6 (C_q) , 158.5 (C_q) , 157.3 (C_q) , 149.1 (C_q) , 136.4 (C_q) , 133.6 (C_q) , 132.4 (CH), 131.5 (CH), 129.9 (C_q), 129.7 (CH), 128.2 (C_q), 126.2 (CH), 126.1 (CH), 125.9 (C_q), 125.5 (CH), 113.7 (CH), 113.1 (CH), 55.2 (CH₃), 55.1 (CH₃), 22.7 (CH₃); IR (ATR) 2937, 2838, 1604, 1510, 1287, 1243, 1170, 1027, 837, 814, 760, 598, 557, 530 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 355 (80) [M⁺], 354 (100), 340 (10), 311 (20), 296 (5), 268 (15), 239 (4), 226 (4), 43 (3); HR-MS (EI) m/zcalcd for C₂₄H₂₁NO₂ 355.1572, found 355.1539. The spectral data are in accordance with those reported in the literature.²³

3,4-Bis(4-fluorophenyl)-1-methylisoquinoline (3ac). The representative procedure was followed using **1a** (68 mg, 0.50 mmol) and 1,2-bis(4-fluorophenyl)acetylene (**2c**) (214 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 8/1) yielded **3ac** (116 mg, 70%) as an orange oil: ¹H NMR (300 MHz, CDCl₃) δ =

8.23–8.15 (m, 1H), 7.65–7.55 (m, 3H), 7.37–7.27 (m, 2H), 7.22–7.12 (m, 2H), 7.11–7.00 (m, 2H), 6.95–6.84 (m, 2H), 3.06 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ = 161.9 (d, ¹J_{C-F} = 247 Hz, C_q), 161.8 (d, ¹J_{C-F} = 247 Hz, C_q), 157.9 (C_q), 148.5 (C_q), 136.8 (d, ⁴J_{C-F} = 3 Hz, C_q), 135.9 (C_q), 133.2 (d, ⁴J_{C-F} = 4 Hz, C_q), 132.8 (d, ³J_{C-F} = 8 Hz, CH), 131.9 (d, ³J_{C-F} = 8 Hz, CH), 130.1 (CH), 128.0 (C_q), 126.6 (CH), 126.1 (C_q), 125.8 (CH), 125.5 (CH), 115.4 (d, ²J_{C-F} = 21 Hz, CH), 114.6 (d, ²J_{C-F} = 21 Hz, CH), 22.8 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = –114.6 (tt, J = 8.8, 5.6 Hz), –115.2 (tt, J = 8.7, 5.5 Hz); IR (ATR) 3035, 1603, 1508, 1390, 1334, 1221, 1154, 1093, 907, 836, 759, 728, 594, 560, 547 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 331 (55) [M⁺], 330 (100), 315 (4), 288 (15), 268 (4); HR-MS (EI) *m*/*z* calcd for C₂₂H₁₅F₂N 331.1173, found 331.1150.

3,4-Diethyl-1-methylisoquinoline (3ad). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 3-hexyne (2d) (82 mg, 1.00 mmol). Purification by column chromatography (nhexane/EtOAc: 12/1) yielded 3ad (86 mg, 86%) as a pale yellow solid (mp 54–57 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.06 (ddd, J = 8.5, 1.3, 0.8 Hz, 1H), 7.96 (dt, J = 8.5, 0.8 Hz, 1H), 7.64 (ddd, J = 8.5, 6.8, 1.3 Hz, 1H), 7.48 (ddd, J = 8.5, 6.8, 1.3 Hz, 1H), 3.03 (q, J = 7.7 Hz, 2H), 2.95 (q, J = 7.7 Hz, 2H), 2.90 (s, 3H), 1.33 (t, J = 7.7 Hz, 3H), 1.27 (t, J = 7.7 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) $\delta = 155.7$ (C₆), 152.5 (C_a), 135.1 (C_a), 129.4 (CH), 127.1 (C_a), 126.1 (CH), 126.0 (C_a), 125.2 (CH), 123.3 (CH), 28.5 (CH₂), 22.3 (CH₃), 20.6 (CH₂), 15.2 (CH₃), 14.9 (CH₃); IR (ATR) 2963, 1614, 1566, 1451, 1391, 1311, 1054, 963, 769, 692, 616 cm⁻¹; MS (EI) m/z (relative intensity) 199 (31) [M⁺], 198 (100), 184 (23), 170 (9), 128 (10), 115 (20), 77 (8), 69 (8); HR-MS (EI) m/z calcd for C₁₄H₁₇N 199.1361, found 199.1355. The spectral data are in accordance with those reported in the literature.¹⁹

1-Methyl-3,4-di-n-propylisoquinoline (3ae). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 4octyne (2e) (110 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 12/1) yielded 3ae (99 mg, 87%) as a yellow oil: ¹H NMR (300 MHz, CDCl₃) δ = 8.06 (dd, J = 8.5, 0.8 Hz, 1H), 7.94 (dd, J = 8.5, 0.8 Hz, 1H), 7.3 (ddd, J = 8.2, 6.9, 1.2 Hz, 1H), 7.47 (ddd, J = 8.2, 6.9, 1.2 Hz, 1H), 3.01-2.92 (m, 2H), 2.92-2.84 (m, 2H), 2.89 (s, 3H), 1.84-1.72 (m, 2H), 1.71-1.58 (m, 2H), 1.07 (t, J = 6.7 Hz, 3H), 1.03 (t, J = 7.1 Hz, 3H); ¹³C NMR (75 MHz, $CDCl_3$) δ = 155.6 (C_a), 151.7 (C_a), 135.4 (C_a), 129.3 (CH), 126.1 (C_q), 126.1 (CH), 126.0 (C_q), 125.2 (CH), 123.5 (CH), 37.4 (CH₂), 29.8 (CH₂), 24.2 (CH₂), 23.8 (CH₂), 22.4 (CH₃), 14.6 (CH₃), 14.4 (CH₃); IR (ATR) 2957, 2870, 1617, 1568, 1454, 1391, 1333, 1027, 754, 614 cm⁻¹; MS (EI) m/z (relative intensity) 227 (40) [M⁺], 212 (80), 198 (100), 184 (50), 171 (55), 128 (23), 115 (16), 77 (6), 43 (31); HR-MS (EI) m/z calcd for C₁₆H₂₁N 227.1674, found 227.1669. The spectral data are in accordance with those reported in the literature.¹⁹

3,4-Di-n-butyl-1-methylisoquinoline (3af). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 5-decyne (2f) (138 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 12/1) yielded 3af (101 mg, 79%) as a yellow oil: ¹H NMR (300 MHz, CDCl₃) δ = 8.05 (dd, J = 8.5, 1.0 Hz, 1H), 7.94 (dd, J = 8.5, 1.0 Hz, 1H), 7.62 (ddd, J = 7.8, 6.8, 1.1 Hz, 1H), 7.46(ddd, J = 7.8, 6.8, 1.1 Hz, 1H), 3.04-2.86 (m, 4H), 2.90 (s, 3H), 1.79–1.65 (m, 2H), 1.65–1.40 (m, 6H), 0.99 (t, J = 7.1 Hz, 3H), 0.97 (t, J = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) $\delta = 155.5$ (C_a), 151.7 (C_q), 135.3 (C_q), 129.3 (CH), 126.1 (C_q), 126.0 (CH), 126.0 (C_a), 125.1 (CH), 123.4 (CH), 35.2 (CH₂), 33.1 (CH₂), 32.8 (CH₂), 27.4 (CH₂), 23.2 (CH₂), 23.0 (CH₂), 22.3 (CH₃), 14.1 (CH₃), 13.9 (CH₃); IR (ATR) 2955, 2858, 1617, 1568, 1504, 1441, 1391, 1336, 1028, 754, 614 cm⁻¹; MS (EI) m/z (relative intensity) 255 (8) [M⁺], 240 (9), 226 (31), 213 (28), 198 (45), 184 (28), 171 (100), 128 (12), 115 (6), 43 (11); HR-MS (EI) m/z calcd for C₁₈H₂₅N 255.1987, found 255.1992.

1,7-Dimethyl-3,4-diphenylisoquinoline (30a). The representative procedure was followed using **10** (75 mg, 0.50 mmol) and diphenylacetylene (**2a**) (178 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 12/1) yielded **30a** (125 mg, 81%) as a pale orange solid (mp 134–139 °C): ¹H NMR (300 MHz,

CDCl₃) δ = 7.96 (dq, *J* = 1.8, 0.9 Hz, 1H), 7.57 (d, *J* = 8.6 Hz, 1H), 7.45–7.30 (m, 6H), 7.26–7.14 (m, 5H), 3.06 (s, 3H), 2.57 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 156.9 (C_q), 148.6 (C_q), 141.0 (C_q), 137.7 (C_q), 136.3 (C_q), 134.1 (C_q), 132.0 (CH), 131.3 (CH), 130.2 (CH), 129.0 (C_q), 128.1 (CH), 127.5 (CH), 127.0 (CH), 126.7 (CH), 126.2 (C_q), 126.0 (CH), 124.4 (CH), 22.7 (CH₃), 21.8 (CH₃); IR (ATR) 3023, 2914, 1551, 1504, 1442, 1386, 1321, 1073, 1027, 831, 767, 755, 696, 567 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 309 (65) [M⁺], 100 (100), 293 (8), 265 (5), 252 (15), 146 (5), 43 (6); HR-MS (ESI) *m*/*z* calcd for C₂₃H₁₉N + H⁺ 310.1590, found 310.1592. The spectral data are in accordance with those reported in the literature.²⁴

5,6-(Methylenedioxy)-1-methyl-3,4-diphenylisoquinoline (3pa). The representative procedure was followed using 1p (100 mg, 0.50 mmol) and diphenylacetylene (2a) (178 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: $10/1 \rightarrow 8/$ $1 \rightarrow 6/1 \rightarrow 2/1$) yielded 3pa (164 mg, 86%) as a white solid (mp 251–254 °C): ¹H NMR (300 MHz, $CDCl_3$) δ = 7.82 (d, J = 8.8 Hz, 1H), 7.37-7.06 (m, 11H), 5.83 (s, 2H), 2.99 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 157.7 (C_q), 150.2 (C_q), 147.6 (C_q), 141.7 (C_q), 140.8 (C_a), 138.4 (C_a), 131.1 (CH), 130.2 (CH), 127.5 (CH), 127.0 (CH), 126.8 (CH), 126.7 (CH), 124.8 (C_q), 123.2 (C_q), 122.5 (C_q), 120.9 (CH), 110.8 (CH), 101.4 (CH₂), 23.4 (CH₃); IR (ATR) 2899, 1626, 1549, 1512, 1432, 1383, 1353, 1279, 1209, 1119, 1049, 891, 794, 760, 744, 698, 644 cm⁻¹; MS (EI) m/z (relative intensity) 339 (100) [M⁺], 338 (98), 310 (18), 292 (14), 278 (9), 267 (6), 239 (6), 176 (5), 139 (9), 77 (7), 43 (8); HR-MS (EI) m/z calcd for C₂₃H₁₇NO₂ 339.1259, found 339.1252

1,4-Dimethyl-3-phenylisoquinoline (3ag). The representative procedure was followed using **1a** (68 mg, 0.50 mmol) and 1-phenyl-1-propyne **(3g)** (116 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 12/1) yielded **3ag** (81 mg, 69%) as a red solid (mp 83–87 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.15 (dd, *J* = 8.5, 0.9 Hz, 1H), 8.04 (dd, *J* = 8.5, 0.9 Hz, 1H), 7.73 (ddd, *J* = 8.5, 6.8, 1.3 Hz, 1H), 7.64–7.54 (m, 3H), 7.52–7.45 (m, 2H), 7.42–7.33 (m, 1H), 2.98 (s, 3H), 2.59 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 155.8 (C_q), 150.6 (C_q), 141.5 (C_q), 136.2 (C_q), 129.8 (CH), 129.8 (CH), 122.1 (C_q), 22.5 (CH₃), 15.4 (CH₃); IR (ATR) 2947, 1683, 1561, 1504, 1437, 1388, 1336, 1159, 1020, 760, 699, 603, 539 cm⁻¹; MS (EI) *m/z* (relative intensity) 233 (50) [M⁺], 232 (100), 217 (10), 202 (4), 189 (7), 128 (5), 115 (8), 77 (6), 43 (11); HR-MS (EI) *m/z* calcd for C₁₇H₁₅N 233.1204, found 233.1186. The spectral data are in accordance with those reported in the literature.¹⁹

4-Ethyl-1-methyl-3-phenylisoquinoline (3ah). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 1phenyl-1-butyne (2h) (130 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 12/1) yielded 3ah (78 mg, 63%) as a pale yellow solid (mp 117-120 °C): ¹H NMR (300 MHz, $CDCl_3$) $\delta = 8.17$ (ddd, J = 8.4, 1.4, 0.7 Hz, 1H), 8.07 (dd, J = 8.6, 1.0 Hz, 1H), 7.73 (ddd, J = 8.4, 6.8, 1.3 Hz, 1H), 7.59 (ddd, J = 8.1, 6.8, 1.2 Hz, 1H), 7.54–7.33 (m, 5H), 2.99 (q, J = 7.5 Hz, 2H), 2.97 (s, 3H), 1.26 (t, J = 7.5 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) $\delta = 155.8$ (C_q) , 150.7 (C_q) , 141.8 (C_q) , 135.1 (C_q) , 129.8 (CH), 129.2 (CH), 128.5 (C_q) , 128.1 (CH), 127.4 (CH), 126.7 (C_q) , 126.3 (CH), 126.1 (CH), 124.1 (CH), 22.5 (CH₃), 21.6 (CH₂), 15.7 (CH₃); IR (ATR) 2963, 1562, 1435, 1390, 1333, 1162, 1027, 861, 771, 748, 685, 620, 590 cm⁻¹; MS (EI) m/z (relative intensity) 247 (54) [M⁺], 246 (100), 231 (21), 217 (10), 202 (6), 189 (5), 128 (5), 115 (7), 77 (6); HR-MS (EI) m/z calcd for C₁₈H₁₇N 247.1361, found 247.1349. The spectral data are in accordance with those reported in the literature.¹⁹

4-*n***-Hexyl-1-methyl-3-phenylisoquinoline (3ai).** The representative procedure was followed using **1a** (68 mg, 0.50 mmol) and 1-phenyl-1-octyne (**2i**) (186 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 12/1) yielded **3ai** (79 mg, 52%) as an orange oil and **3ai'** (9 mg, 6%) as an orange oil: ¹H NMR (300 MHz, CDCl₃) δ = 8.15 (ddd, *J* = 8.4, 1.3, 0.8 Hz, 1H), 8.05 (dd, *J* = 8.4, 0.8 Hz, 1H), 7.72 (ddd, *J* = 8.4, 6.8, 1.3 Hz, 1H), 7.58 (ddd, *J* = 8.4, 6.8, 1.3 Hz, 1H), 7.58 (ddd, *J* = 8.4, 6.8, 1.3 Hz, 1H), 7.58 (ddd, *J* = 8.4, 6.8, 1.3 Hz, 1H), 7.59 (s, 3H), 1.70–1.56 (m, 2H), 1.38–1.14 (m, 6H), 0.84 (t, *J* = 6.7 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 155.7 (C_a), 150.9 (C_a), 141.9 (C_a),

135.4 (C_q), 129.7 (CH), 129.3 (CH), 128.1 (CH), 127.4 (C_q), 127.3 (CH), 126.6 (C_q), 126.2 (CH), 126.1 (CH), 124.2 (CH), 31.4 (CH₂), 31.2 (CH₂), 29.5 (CH₂), 28.5 (CH₂), 22.5 (CH₂), 22.5 (CH₃), 14.0 (CH₃); IR (ATR) 2924, 2856, 1614, 1562, 1504, 1436, 1391, 1333, 1029, 755, 698, 616, 592 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 303 (35) [M⁺], 260 (10), 246 (50), 217 (15), 189 (5); HR-MS (EI) *m*/*z* calcd for $C_{22}H_{25}N$ 303.1987, found 303.1982.

3-*n***-Hexyl-1-methyl-4-phenylisoquinoline (3ai').** Data: ¹H NMR (300 MHz, CDCl₃) δ = 8.14–8.07 (m, 1H), 7.53–7.38 (m, SH), 7.37–7.30 (m, 1H), 7.30–7.22 (m, 2H), 2.99 (s, 3H), 2.69–2.58 (m, 2H), 1.77–1.55 (m, 2H), 1.28–1.08 (m, 6H), 0.79 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 157.4 (C_q), 151.7 (C_q), 137.9 (C_q), 136.0 (C_q), 130.4 (CH), 129.5 (CH), 128.9 (C_q), 128.3 (CH), 127.3 (CH), 125.8 (CH), 125.6 (CH), 125.4 (C_q), 125.3 (CH), 35.7 (CH₂), 31.6 (CH₂), 30.4 (CH₂), 29.3 (CH₂), 22.5 (CH₂), 22.5 (CH₃), 14.0 (CH₃); IR (ATR) 2925, 2855, 1713, 1563, 1505, 1442, 1392, 1336, 1028, 758, 699, 626, 599 cm⁻¹; MS (EI) *m/z* (relative intensity) 303 (4) [M⁺], 288 (4), 274 (5), 260 (15), 246 (20), 232 (100), 217 (10), 189 (8), 43 (7); HR-MS (EI) *m/z* calcd for C₂₂H₂₅N 303.1987, found 303.1976.

1-{4-(1,4-Dimethylisoquinolin-3-yl)phenyl}ethanone (3aj). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 1-{4-(prop-1-yn-1-yl)phenyl}ethanone (2j) (158 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 2/ 1) yielded **3aj** (97 mg, 70%) as a yellow solid (mp 149–153 °C): ¹H NMR (300 MHz, $CDCl_3$) δ = 8.17 (ddd, J = 8.3, 1.3, 0.7 Hz, 1H), 8.09-8.04 (m, 1H), 8.07 (d, J = 8.5 Hz, 2H), 7.76 (ddd, J = 8.3, 6.9, 1.3 Hz, 1H), 7.69 (d, J = 8.5 Hz, 2H), 7.63 (ddd, J = 8.2, 6.9, 1.2 Hz, 1H), 2.98 (s, 3H), 2.66 (s, 3H), 2.60 (s, 3H); $^{13}\mathrm{C}$ NMR (75 MHz, $CDCl_3$) $\delta = 197.9 (C_q)$, 156.2 (C_q), 149.3 (C_q), 146.3 (C_q), 136.1 (C_q), 135.9 (C_q), 130.2 (CH), 130.0 (CH), 128.1 (CH), 126.7 (CH), 126.3 (C_q), 126.1 (CH), 124.1 (CH), 122.6 (C_q), 26.7 (CH₃), 22.5 (CH₃), 15.3 (CH₃); IR (ATR) 2920, 1681, 1605, 1565, 1261, 1012, 957, 853, 834, 762, 733, 599, 540 cm⁻¹; MS (EI) m/z (relative intensity) 275 (59) [M⁺], 274 (100), 231 (27), 217 (8), 188 (6), 129 (5), 115 (5), 43 (13); HR-MS (EI) m/z calcd for $C_{19}H_{17}NO$ 275.1310, found 275.1299.

4-n-Butyl-1-methyl-3-(p-tolyl)isoquinoline (3ak). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 1-(p-tolyl)-1-hexyne (2k) (172 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 12/1) yielded 3ak (98 mg, 68%) as an orange oil and 3ak' (11 mg, 8%) as an orange oil: ¹H NMR (300 MHz, CDCl₃) δ = 8.14 (dd, J = 8.6, 0.9 Hz, 1H), 8.04 (dt, J = 8.6, 0.9 Hz, 1H), 7.70 (ddd, J = 8.3, 6.8, 1.3 Hz, 1H), 7.56 (ddd, J = 8.3, 6.8, 1.2 Hz, 1H), 7.39 (d, J = 8.1 Hz, 2H), 7.25 (d, J = 8.1 Hz, 2H), 3.03-2.91 (m, 2H), 2.95 (s, 3H), 2.41 (s, 3H), 1.70-1.56 (m, 2H), 1.34 (dt, J = 7.3, 7.3 Hz, 2H), 0.86 (t, J = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 155.6 (C_q), 150.9 (C_q), 139.0 (C_q), 136.9 (C_q), 135.4 (C_q), 129.6 (CH), 129.1 (CH), 128.7 (CH), 127.2 (C_q), 126.5 (C_q), 126.2 (CH), 126.0 (CH), 124.2 (CH), 33.4 (CH₂), 28.3 (CH₂), 23.0 (CH₂), 22.5 (CH₃), 21.2 (CH₃), 13.8 (CH₃); IR (ATR) 2955, 2923, 2869, 1614, 1563, 1513, 1438, 1391, 1333, 1026, 825, 755, 726, 615, 587, 539, 497 cm⁻¹; MS (EI) m/z (relative intensity) 289 (50) [M⁺], 260 (70), 246 (100), 231 (30), 216 (8), 202 (10), 115 (5), 43 (11); HR-MS (EI) m/z calcd for C₂₁H₂₃N 289.1830, found 289.1833.

3-*n***-Butyl-1-methyl-4-(***p***-tolyl)isoquinoline (3ak').** Data: ¹H NMR (300 MHz, CDCl₃) δ = 8.13–8.05 (m, 1H), 7.51–7.43 (m, 2H), 7.39–7.32 (m, 1H), 7.28 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 2.98 (s, 3H), 2.70–2.61 (m, 2H), 2.45 (s, 3H), 1.70–1.55 (m, 2H), 1.31–1.16 (m, 2H), 0.78 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 157.2 (C_q), 151.8 (C_q), 136.8 (C_q), 136.2 (C_q), 134.8 (C_q), 130.3 (CH), 129.4 (CH), 129.0 (CH), 128.9 (C_q), 125.9 (CH), 125.6 (CH), 125.4 (C_q), 125.3 (CH), 35.5 (CH₂), 32.7 (CH₂), 22.7 (CH₂), 22.5 (CH₃), 21.3 (CH₃), 13.9 (CH₃); IR (ATR) 2955, 2924, 1614, 1563, 1517, 1439, 1391, 1336, 1107, 1024, 965, 817, 758, 613 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 289 (5) [M⁺], 274 (10), 260 (15), 246 (100), 231 (15), 216 (5), 202 (8), 189 (6), 122 (5), 43 (12); HR-MS (EI) *m*/*z* calcd for C₂₁H₂₃N 289.1830, found 289.1822. **3-([1,1'-Biphenyl]-4-***n***-butyl-1-methylisoquinoline** 0.50 mmol) and 1,1'-biphenyl-1-hexyne (2l) (234 mg, 1.00 mmol). Purification by column chromatography (*n*-hexane/EtOAc: 12/1) yielded **3al** (90 mg, 51%) as a pale yellow solid (mp 129–130 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.17 (d, *J* = 8.4 Hz, 1H), 8.07 (d, *J* = 8.4 Hz, 1H), 7.78–7.55 (m, 8H), 7.51–7.43 (m, 2H), 7.41–7.33 (m, 1H), 3.10–2.96 (m, 2H), 2.99 (s, 3H), 1.76–1.60 (m, 2H), 1.38 (dt, *J* = 7.4, 7.4 Hz, 2H), 0.88 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 155.9 (C_q), 150.5 (C_q), 141.1 (C_q), 140.9 (C_q), 140.1 (C_q), 135.4 (C_q), 129.7 (CH), 129.7 (CH), 128.7 (CH), 127.4 (C_q), 127.2 (CH), 127.1 (CH), 126.9 (CH₂), 23.0 (CH₂), 22.5 (CH₃), 13.8 (CH₃); IR (ATR) 2926, 2870, 1563, 1436, 1387, 1334, 1006, 927, 814, 761, 732, 692, 598, 485 cm⁻¹; MS (EI) *m*/*z* (relative intensity) 351 (25) [M⁺], 322 (45), 308 (100), 231 (6), 77 (10); HR-MS (EI) *m*/*z* calcd for C₂₆H₂₅N 351.1987, found 351.1979.

4-n-Butyl-1-methyl-3-(4-methoxyphenyl)isoquinoline (3am). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 1-(4-methoxyphenyl)-1-hexyne (2m) (188 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 12/1) yielded 3am (87 mg, 57%) as a red oil: ¹H NMR (300 MHz, $CDCl_3$) δ = 8.14 (ddd, J = 8.4, 1.3, 0.7 Hz, 1H), 8.03 (dd, J = 8.6, 0.7 Hz, 1H), 7.70 (ddd, J = 8.2, 6.8, 1.3 Hz, 1H), 7.56 (ddd, J = 8.2, 6.8, 1.1 Hz, 1H), 7.44 (d, J = 8.8 Hz, 2H), 6.99 (d, J = 8.8 Hz, 2H), 3.86 (s, 3H), 3.06-2.90 (m, 2H), 2.95 (s, 3H), 1.71-1.54 (m, 2H), 1.35 (dd, J = 7.3 Hz, 2H), 0.86 (t, J = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 158.9 (C_q), 155.6 (C_q), 150.5 (C_q), 135.4 (C_q), 134.4 (C_q), 130.5 (CH), 129.6 (CH), 127.2 (C_q), 126.5 (C_q), 126.2 (CH), 125.9 (CH), 124.1 (CH), 113.5 (CH), 55.3 (CH₃), 33.4 (CH₂), 28.3 (CH₂), 22.9 (CH₂), 22.5 (CH₃), 13.8 (CH₃); IR (ATR) 2957, 2926, 2868, 1609, 1567, 1512, 1463, 1436, 1391, 1376, 1291, 1250, 1176, 1021, 836, 753, 615, 588, 574, 541 cm⁻¹; MS (EI) m/z (relative intensity) 305 (72) [M⁺], 276 (71), 262 (100), 247 (35), 230 (6), 218 (23), 43 (8); HR-MS (EI) m/z calcd for $C_{21}H_{23}NO$ 305.1780, found 305.1771.

4-n-Butyl-3-(4-fluorophenyl)-1-methylisoquinoline (3an). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 1-(4-fluoroyphenyl)-1-hexyne (2n) (176 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 12/1) yielded 3an (88 mg, 60%) as a pale brown solid (mp 58-60 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.15 (ddd, J = 8.4, 1.3, 0.7 Hz, 1H), 8.04 (dd, *J* = 8.4, 0.7 Hz, 1H), 7.72 (ddd, *J* = 8.4, 6.8, 1.3 Hz, 1H), 7.58 (ddd, J = 8.4, 6.8, 1.3 Hz, 1H), 7.51-7.41 (m, 2H), 7.20-7.07 (m, 2H), 3.01-2.89 (m, 2H), 2.95 (s, 3H), 1.67-1.53 (m, 2H), 1.40-1.25 (m, 2H), 0.84 (t, J = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) $\delta =$ 162.3 (d, ${}^{1}J_{C-F}$ = 246 Hz, C_a), 155.9 (C_a), 149.8 (C_a), 137.9 (d, ${}^{4}J_{C-F}$ = 3 Hz, C_q), 135.3 (C_q), 131.0 (d, ${}^{3}J_{C-F}$ = 8 Hz, CH), 129.9 (CH), 127.5 (C_a), 126.7 (C_a), 126.3 (CH), 126.3 (CH), 124.2 (CH), 115.0 $(d, {}^{2}J_{C-F} = 21 \text{ Hz}, \text{ CH}), 33.3 (CH_{2}), 28.2 (CH_{2}), 22.9 (CH_{2}), 22.5$ (CH_3) , 13.7 (CH_3) ; ¹⁹F NMR (282 MHz, CDCl₃) $\delta = -115.3$ (tt, J =8.8, 5.5 Hz); IR (ATR) 2931, 2868, 1603, 1434, 1387, 1335, 1218, 1091, 837, 761, 727, 711, 616 cm⁻¹; MS (EI) *m/z* (relative intensity) 293 (40) [M⁺], 264 (36), 250 (100), 235 (15), 220 (4), 207 (4), 147 (4); HR-MS (EI) m/z calcd for C₂₀H₂₀FN 293.1580, found 293.1584.

Ethyl 4-(4-n-butyl-1-methylisoquinolin-3-yl)benzoate (3ao). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and ethyl 4-(hex-1-yn-1-yl)benzoate (20) (230 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 12/1) yielded 3ao (81 mg, 47%) as a red oil: ¹H NMR (300 MHz, $CDCl_3$) $\delta = 8.15$ (dd, J = 8.6, 0.9 Hz, 1H) 8.13 (d, J = 8.1 Hz, 2H), 8.04 (dd, J = 8.6, 0.9 Hz, 1H), 7.73 (ddd, J = 8.4, 6.9, 1.4 Hz, 1H), 7.64–7.54 (m, 3H), 4.40 (q, J = 7.1 Hz, 2H), 3.00–2.86 (m, 2H), 2.95 (s, 3H), 1.65–1.53 (m, 2H), 1.41 (t, J = 7.1 Hz, 3H), 1.30 (dt, J = 7.4, 7.4 Hz, 2H), 0.82 (t, J = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) $\delta =$ 166.6 (C_q), 156.1 (C_q), 149.7 (C_q), 146.4 (C_q), 135.2 (C_q), 129.9 (CH), 129.4 (CH), 129.4 (CH), 129.3 (C_q), 127.6 (C_q), 126.8 (C_q), 126.5 (CH), 126.2 (CH), 124.2 (CH), 60.9 (CH₂), 33.4 (CH₂), 28.2 (CH₂), 22.9 (CH₂), 22.5 (CH₃), 14.3 (CH₃), 13.7 (CH₃); IR (ATR) 2957, 2871, 1713, 1611, 1563, 1508, 1391, 1366, 1267, 1174, 1098, 1019, 866, 757, 712, 616 cm⁻¹; MS (EI) m/z (relative intensity) 347 (64) [M⁺], 318 (100), 304 (15), 290 (14), 276 (8), 260 (25), 244

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(11), 231 (61), 216 (11), 202 (10), 189 (5), 115 (5), 43 (18); HR-MS (EI) *m*/*z* calcd for C₂₃H₂₅NO₂ 347.1885, found 347.1870.

4-(4-Hydroxy-n-butyl)-1-methyl-3-phenylisoquinoline (3ap). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 6-phenylhex-5-yn-1-ol (2p) (174 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 2/1) yielded 3ap (106 mg, 73%) a pale brown solid (mp 96-101 °C): ¹H NMR (300 MHz, CDCl₃) δ = 8.17 (dd, J = 8.5, 1.3, 0.8 Hz, 1H), 8.06 (dd, J = 8.5, 0.8 Hz, 1H), 7.73 (ddd, J = 8.5, 6.8, 1.3 Hz, 1H), 7.59 (ddd, J = 8.1, 6.8, 1.2 Hz, 1H), 7.52-7.34 (m, 5H), 3.48 (t, J = 6.5 Hz, 2H), 3.06–2.90 (m, 2H), 2.97 (s, 3H), 1.75–45 (m, 5H); ¹³C NMR $(75 \text{ MHz}, \text{CDCl}_3) \delta = 156.0 (C_q), 151.0 (C_q), 141.7 (C_q), 135.3 (C_q),$ 130.0 (CH), 129.3 (CH), 128.1 (CH), 127.4 (CH), 126.9 (C₀), 126.6 (C_a), 126.3 (CH), 126.2 (CH), 124.1 (CH), 62.2 (CH₂), 32.5 (CH₂), 29.0 (CH₂), 27.2 (CH₂), 22.4 (CH₃); IR (ATR) 3273, 2936, 2863, 1615, 1564, 1444, 1395, 1335, 1141, 1053, 1030, 982, 759, 707, 628 cm⁻¹; MS (EI) m/z (relative intensity) 291 (50) [M⁺], 246 (68), 232 (100), 217 (14), 202 (6), 189 (7), 115 (5), 77 (5), 43 (6); HR-MS (EI) *m/z* calcd for C₂₀H₂₁NO 291.1623, found 291.1623.

4-(4-Chloro-n-butyl)-1-methyl-3-phenylisoquinoline (3aq). The representative procedure was followed using 1a (68 mg, 0.50 mmol) and 6-chloro-1-phenylhexyne (2q) (193 mg, 1.00 mmol). Purification by column chromatography (n-hexane/EtOAc: 12/1) yielded 3aq (104 mg, 67%) as a yellow oil: ¹H NMR (300 MHz, $CDCl_3$) $\delta = 8.16$ (ddd, I = 8.4, 1.3, 0.8 Hz, 1H), 8.04 (dd, I = 8.4, 0.8Hz, 1H), 7.73 (ddd, J = 8.4, 6.9, 1.3 Hz, 1H), 7.59 (ddd, J = 8.4, 6.9, 1.3 Hz, 1H), 7.53–7.34 (m, 5H), 3.42 (t, J = 6.2 Hz, 2H), 3.03–2.95 (m, 2H), 2.97 (s, 3H), 1.86-1.68 (m, 4H); ¹³C NMR (75 MHz, $CDCl_3$) δ = 156.1 (C_q), 151.1 (C_q), 141.6 (C_q), 135.2 (C_q), 129.9 (CH), 129.2 (CH), 128.2 (CH), 127.5 (CH), 126.6 (C_q), 126.4 (C_q), 126.3 (CH), 126.2 (CH), 124.0 (CH), 44.4 (CH₂), 32.3 (CH₂), 28.1 (CH₂), 27.5 (CH₂), 22.5 (CH₃); IR (ATR) 2953, 1614, 1561, 1504, 1437, 1391, 1331, 1027, 756, 699, 648, 616, 592 cm⁻¹; MS (EI) m/z(relative intensity) 309 (41) [M⁺], 246 (95), 232 (100), 217 (16), 202 (6), 189 (6), 115 (6), 77 (5); HR-MS (EI) m/z calcd for C₂₀H₂₀ClN 309.1284, found 309.1297.

Competition Experiment with Alkynes 2 (Scheme 5). A mixture of acetophenone oxime (1a) (68 mg, 0.50 mmol), diphenylacetylene (2a) (356 mg, 2.00 mmol), 4-octyne (2e) (220 mg, 2.00 mmol), $[RuCl_2(p\text{-cymene})]_2$ (15.3 mg, 0.025 mmol, 5.0 mol %), and KPF₆ (28 mg, 0.15 mmol, 30 mol %) in MeOH (2.00 mL) was stirred at 60 °C for 24 h. At ambient temperature, EtOAc (15 mL) was added, and the solvents were removed in vacuo. Purification of the remaining residue by column chromatography (*n*-hexane/EtOAc: 12/1) yielded **2aa** (80 mg, 54%) as a white solid and **2ae** (36 mg, 32%) as a yellow oil.

Competition Experiment with Oximes 1c and 1q (Scheme 6a). A mixture of oxime 1c (118 mg, 0.55 mmol), oxime 1q (91 mg, 0.55 mmol), diphenylacetylene (2a) (89 mg, 0.5 mmol), $[RuCl_2(p-cymene)]_2$ (15.3 mg, 0.025 mmol, 5.0 mol %), and KPF₆ (28 mg, 0.15 mmol, 30 mol %) in MeOH (2.00 mL) was stirred at 60 °C for 24 h. At ambient temperature, EtOAc (15 mL) was added, and the solvents were removed in vacuo. Purification of the remaining residue by column chromatography (*n*-hexane/EtOAc: 12/1) yielded 3ca (8 mg, 5%) as an orange solid and 3qa (62 mg, 38%) as a pale yellow solid (mp 175–177 °C).

6-Methoxy-1-methyl-3,4-diphenylisoquinoline (3qa). Data: ¹H NMR (300 MHz, CDCl₃) δ = 8.09 (d, *J* = 9.1 Hz, 1H), 7.38– 7.28 (m, 5H), 7.25–7.13 (m, 6H), 6.91 (d, *J* = 2.6 Hz, 1H), 3.71 (s, 3H), 3.01 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ = 160.5 (C_q), 156.9 (C_q), 150.1 (C_q), 141.2 (C_q), 138.0 (C_q), 137.8 (C_q), 131.2 (CH), 130.2 (CH), 128.5 (C_q), 128.2 (CH), 127.5 (CH), 127.4 (CH), 127.0 (CH), 126.8 (CH), 121.8 (C_q), 118.6 (CH), 104.4 (CH), 55.2 (CH₃), 22.6 (CH₃); IR (ATR) 2923, 1618, 1500, 1410, 1273, 1229, 1205, 1070, 1024, 853, 823, 767, 696, 611 cm⁻¹; MS (EI) *m/z* (relative intensity) 325 (55) [M⁺], 324 (100), 281 (32), 239 (6), 139 (5), 43 (10); HR-MS (EI) *m/z* calcd for C₂₃H₁₉NO 325.1467, found 325.1471. The spectral data are in accordance with those reported in the literature.¹⁹ Competition Experiment with Oximes 1c and 1b (Scheme 6b). A mixture of oxime 1c (118 mg, 0.55 mmol), oxime 1b (82 mg, 0.55 mmol), diphenylacetylene (2a) (89 mg, 0.5 mmol), $[RuCl_2(p-cymene)]_2$ (15.3 mg, 0.025 mmol, 5.0 mol %), and KPF₆ (28 mg, 0.15 mmol, 30 mol %) in MeOH (2.00 mL) was stirred at 60 °C for 24 h. At ambient temperature, EtOAc (15 mL) was added, and the solvents were removed in vacuo. Purification of the remaining residue by column chromatography (*n*-hexane/EtOAc: 12/1) yielded 3ca (13 mg, 7%) as an orange solid and 3ba (45 mg, 29%) as a pale orange solid.

Reaction of Oxime 1a in MeOH- d_4 (Scheme 7a). A mixture of acetophenone oxime (1a) (68 mg, 0.50 mmol), $[RuCl_2(p-cymene)]_2$ (15.3 mg, 0.025 mmol, 5.0 mol %), and KPF₆ (28 mg, 0.15 mmol, 30 mol %) in MeOH- d_4 (2.00 mL) was stirred at 60 °C for 24 h. At ambient temperature, EtOAc (50 mL) and H₂O (20 mL) were added, and the organic layer was separated. The aqueous layer was extracted with EtOAc (50 mL), and the combined organic layers were dried over Na₂SO₄. Evaporation of the solvents yielded 1a (62 mg, 92%) with less than 5% deuterium incorporation in the *ortho*-position estimated by ¹H NMR spectroscopy.

Reaction of Substrates 1a and 2e in MeOH- d_4 (Scheme 7b). A mixture of acetophenone oxime (1a) (68 mg, 0.50 mmol), 4-octyne (2e) (110 mg, 1.00 mmol), $[\text{RuCl}_2(p\text{-cymene})]_2$ (15.3 mg, 0.025 mmol, 5.0 mol %), and KPF₆ (28 mg, 0.15 mmol, 30 mol %) in MeOH- d_4 (2.00 mL) was stirred at 60 °C for 24 h. At ambient temperature, EtOAc (15 mL) was added, and the solvents were removed in vacuo. Purification of the remaining residue by column chromatography (*n*-hexane/EtOAc: 12/1) yielded **3ae** (86 mg, 76%) with 15% deuterium incorporation in the *ortho*-position and 57% deuterium incorporation at the Methyl-Group as estimated by ¹H NMR spectroscopy.

Reaction of Substrates 1a-d₅ and 2e (Scheme 7c). A mixture of acetophenone oxime- d_5 (1a- d_5) (70 mg, 0.50 mmol), 4-octyne (2e) (110 mg, 1.00 mmol), [RuCl₂(*p*-cymene)]₂ (15.3 mg, 0.025 mmol, 5.0 mol %), and KPF₆ (28 mg, 0.15 mmol, 30 mol %) in MeOH (2.00 mL) was stirred at 60 °C for 24 h. At ambient temperature, EtOAc (15 mL) was added, and the solvents were removed in vacuo. Purification of the remaining residue by column chromatography (n-hexane/ EtOAc: 12/1) yielded **3ae**- d_4 (107 mg, 91%) as a yellow oil with 16% hydrogen incorporation in the ortho-position as estimated by ¹H NMR spectroscopy: ¹H NMR (300 MHz, CDCl₃) δ = 3.07–2.81 (m, 4H), 2.89 (s, 3H), 1.86–1.72 (m, 2H) 1.73–1.59 (m, 2H), 1.08 (t, J = 7.3Hz, 3H), 1.04 (t, J = 7.3 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃) $\delta =$ 155.5 (C_a), 151.5 (C_a), 135.2 (C_q), 128.8 (t, J = 24 Hz, CD), 126.0 (C_q) , 125.8 (C_q) , 125.2 (t, J = 24 Hz, CD), 124.6 (t, J = 24 Hz, CD), 123.1 (t, J = 24 Hz, CD), 37.4 (CH₂), 29.7 (CH₂), 24.1 (CH₂), 23.8 (CH₂), 22.3 (CH₃), 14.5 (CH₃), 14.3 (CH₃); IR (ATR) 2957, 2930, 2871, 1593, 1555, 1451, 1402, 1376, 1307, 1275, 1089, 878, 647, 559 cm^{-1} ; MS (EI) m/z (relative intensity) 231 (65) [M⁺], 216 (100), 201 (82), 184 (73), 175 (88), 160 (23), 145 (13), 131 (33), 118 (20), 92 (10), 79 (10), 41 (10); HR-MS (ESI) m/z calcd for $C_{16}H_{17}D_4N + H^+$ 232.1998, found 232.1998.

ASSOCIATED CONTENT

S Supporting Information

Copies of 1 H and 13 C NMR spectra for all products. This material is available free of charge via the Internet at http:// pubs.acs.org.

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Notes

The authors declare no competing financial interest.

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